# TSK-GEL® Aminopak Products

**Part Numbers:** 13181, 12cm x 4.6mm ID

NOTE: No guard column products are offered to protect TSK-GEL Aminopak columns. It is recommended to use a frit or

screen fiter (<2 micron porosity) between the injector and the column. Samples and mobile phases are best

filtered and degassed through a 0.45 micron membrane filter.

This sheet contains the recommended operating conditions and the specifications for TSK-GEL Aminopak columns. Aminopak columns contain spherical, porous, 5 micron particles that are functionalized with strong cation exchange groups balanced by sodium counter-ions. Installation instructions and column care information are described in a separate Instruction Manual.

### A. OPERATING CONDITIONS

1. Shipping Solvent: Citric Acid Buffer (Buffer 2 for the Ninhydrin Method; see back page)

2. Max. Flow Rate: 0.5 ml/min at 55°C

When a buffer with high viscosity is used, the maximum flow rate may have to be reduced so as not to exceed the

maximum pressure drop.

3. Standard Flow Rate: 0.3 - 0.4 ml/min

Max. Pressure: 80 kg/cm<sup>2</sup> = 1200 psi

pH Range: 2.0 - 14.0
 Salt Conc.: <1.5 Molar</li>
 Organic Conc.: <20%</li>

8. Temperature: 25 - 80°C, during normal operation of the column; 15-30°C, during column storage.

9. Cleaning Solvents: 0.2M NaOH. We recommend to flush the column with 2ml of 0.2M NaOH after each amino acid analysis.

10. Storage: For overnight storage, flush the column with the starting gradient buffer at 0.1 ml/min. Store the column in the

shipping solvent when it will not be used the next day. Prevent air from entering the column!

11. Amino Acid Analysis: Turn this page over for the recommended buffer compositions for separating free amino acids followed by

post-column reaction with ninhydrin or o-phthalaldehyde (OPA) and UV-Vis and fluorescence detection.

#### B. SPECIFICATIONS

The performance of TSK-GEL Aminopak columns is tested under the conditions described in the Data Sheet. All columns have passed the following quality control specifications:

Number of Theoretical Plates >1,250 (N):
 Asymmetry Factor (AF): 1.0 - 1.35

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## **Analyzing Free Amino Acids on TSKgel Aminopak**

### Ninhydrin Method

| BUFFER 1 | Sodium citrate ( $C_6H_8O_7Na_3.2H_20$ ) citric acid ( $C_6H_8O_7.H_20$ ) ethanol β-thiodiglycol ( $C_4H_{10}O_2S$ ) 30% Brij-35 (polyoxyethylene laurylether) n-caprylic acid (octanoic acid) Adjust the pH to 3.26 with HCl. | 19.6<br>28.0<br>80<br>5.0<br>3.3<br>0.1 | g/l<br>g/l<br>ml/l<br>ml/l<br>ml/l |
|----------|--|---|------------------------------------|
| BUFFER 2 | Sodium citrate citric acid β-thiodiglycol 30% Brij-35 n-caprylic acid (C <sub>8</sub> H <sub>16</sub> O <sub>2</sub> ) Adjust the pH to 4.30 with HCl.   | 19.6<br>14.0<br>5.0<br>3.3<br>0.1       | g/l<br>g/l<br>ml/l<br>ml/l<br>ml/l |
| BUFFER 3 | Sodium citrate Sodium borate (Na <sub>2</sub> B <sub>4</sub> O <sub>7</sub> .10H <sub>2</sub> 0) Sodium chloride 30% Brij-35 n-caprylic acid Adjust the pH to 9.40 with HCI.   | 14.7<br>9.5<br>35.1<br>3.3<br>0.1       | g/l<br>g/l<br>g/l<br>ml/l<br>ml/l  |
| CLEANING | 0.2M NaOH  | 8.0                                     | g/l                                |

SOLVENT EXCHANGE Buffer 1 from 0 to 11 min; buffer 2 to 22 min., buffer to 3 to 45 min., followed by 5 min clean-up solution and return to buffer 1 at 50 minutes.

### **OPA Method**

| BUFFER 1 | Sodium citrate ethanol n-caprylic acid Adjust the pH to 3.26 with HCl.                       | 19.6<br>80<br>0.1          | g/l<br>ml/l<br>ml/l       |
|----------|--|----------------------------|---------------------------|
| BUFFER 2 | Sodium citrate<br>n-caprylic acid<br>Adjust the pH to 4.30 with HCl.                         | 19.6<br>0.1                | g/l<br>ml/l               |
| BUFFER 3 | Sodium citrate Sodium borate Sodium chloride n-caprylic acid Adjust the pH to 9.40 with HCl. | 14.7<br>9.5<br>35.1<br>0.1 | g/l<br>g/l<br>g/l<br>ml/l |
| CLEANING | 0.2M NaOH<br>n-caprylic acid   | 8.0<br>0.1                 | g/l<br>ml/l               |

SOLVENT EXCHANGE Buffer 1 from 0 to 11 min; buffer 2 to 22 min., buffer 3 to 44 min., followed by 11 min clean-up solution and return to buffer 1 at 55 minutes.