



# A Novel Desalting Column With High Mechanical Strength for Faster Desalting of Proteins

**Atis Chakrabarti and Roy Eksteen**

Tosoh Bioscience LLC, King of Prussia, PA 19406



# Introduction

- Desalting is a process to remove or reduce salt from a liquid stream.
- Proteins elute at high or elevated salt concentration in such chromatographic modes as hydrophobic interaction (HIC), ion exchange (IEC) and size exclusion chromatography (SEC).
- SEC mobile phases for protein analysis may also contain denaturants such as guanidine hydrochloride and urea in addition to salt and buffer.
- Desalting on the basis of size exclusion chromatography is widely used in biochemical purifications.
- Desalting and buffer exchange of proteins or polynucleotides can also be performed by dialysis, ultra filtration, or by using spin-columns.



# Introduction

- Desalting columns are characterized by a low exclusion limit and a large pore volume.
- Salts can fully access all pores, while proteins and other high MW species are excluded from the pores and elute in the void volume as a narrow concentrated peak.
- Columns packed with conventional packing materials such as dextran, cellulose and polyacrylamide have limited physical stability and are not suitable when fast desalting is desired.
- We increased the mechanical strength of polyacrylamide gel by four-fold over that of conventional gels
- TSKgel BioAssist DS columns contain 15 $\mu$ m particles packed in 4.6mm ID x 15cm and 10mm ID x 15cm PEEK columns.

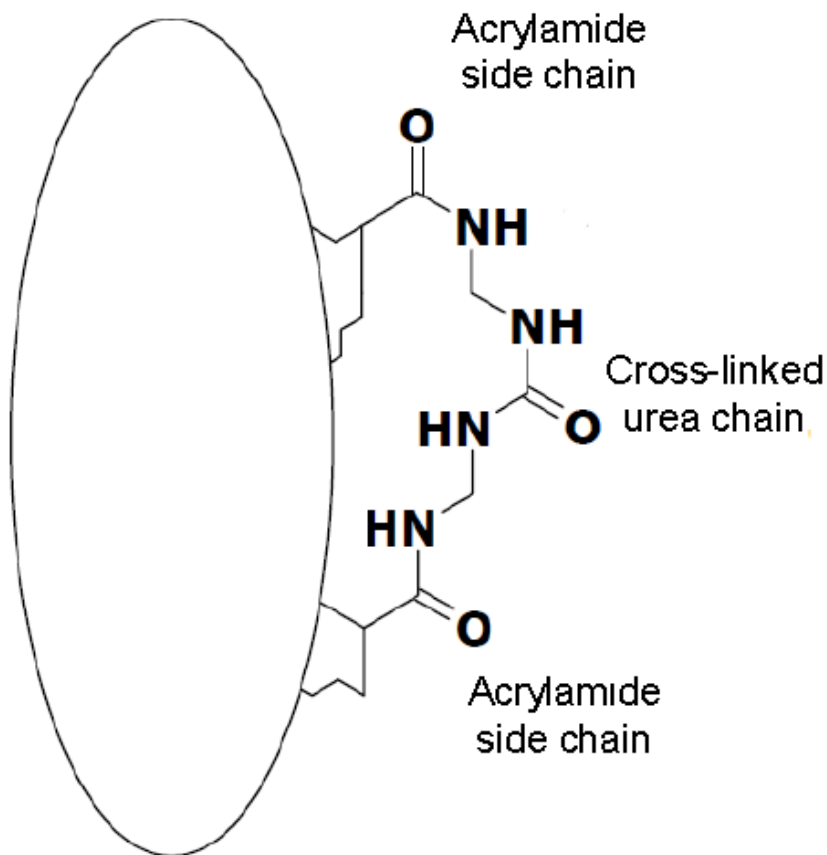


# Objective

To show the usefulness of the new TSKgel BioAssist DS columns for efficient desalting using a conventional HPLC system.



# Chemical structure of polyacrylamide beads\*



- Backbone of beads is based on polyacrylamide
- Side chains cross-linked with one another through urea

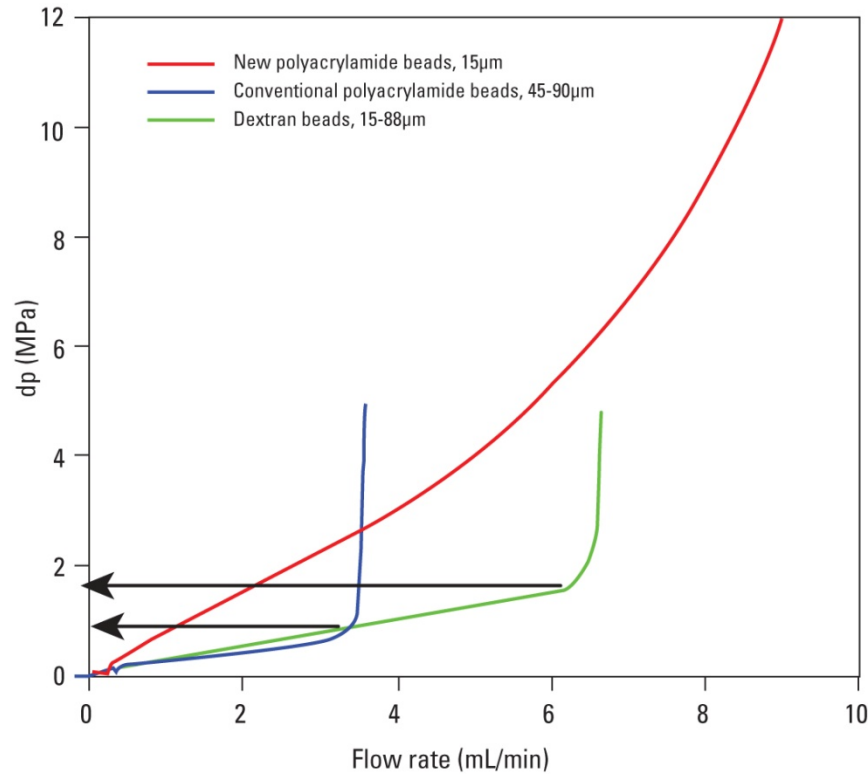


- Highly cross-linked polyacrylamide beads display high mechanical strength and low hydrophobicity

\*US Patent No. 7,659,348



# Mechanical strength

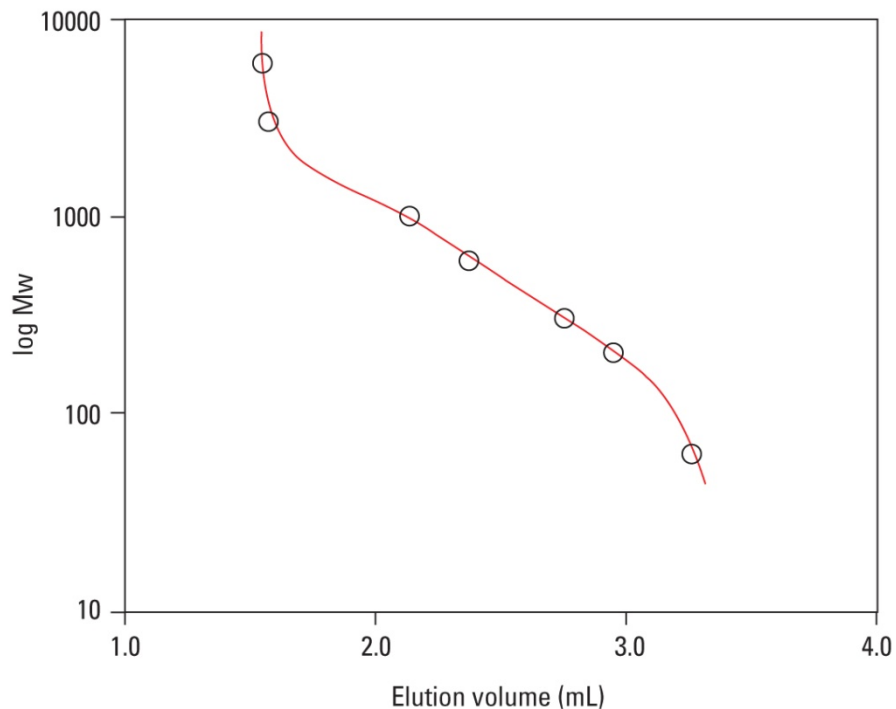


Column size: 4.6mm ID x 10cm  
Mobile phase: D-H<sub>2</sub>O

- **Conventional beads collapsed at pressures below 1.6MPa (< 250psi).**
- **TSKgel BioAssist DS polyacrylamide beads did not collapse at 12MPa (1750psi).**



# Calibration curve for new polyacrylamide beads



Column size: 6.0mm ID x 15cm  
Mobile phase: DI H<sub>2</sub>O  
Flow rate: 0.5mL/min  
Sample: ethylene glycol, PEG

**Exclusion limit PEG 2500 MW**



# Characteristics of TSKgel BioAssist DS Desalting columns

- Packing material: Urea cross-linked polyacrylamide
- Particle Diameter: 15 $\mu$ m (Uniform)
- Pore Size excludes: ca. 2500 MW PEG
- Particle porosity: ca. 60%
- Maximum pressure: 4Mpa (< 600psi)





# Material and methods: Chromatographic conditions (size exclusion experiment)

- Column: TSKgel G3000SW<sub>XL</sub>, 5 $\mu$ m, 7.8mm ID x 30cm (S1237-08R)
- Mobile Phase: 100mmol/L KH<sub>2</sub>PO<sub>4</sub>/K<sub>2</sub>HPO<sub>4</sub>, pH 6.7, 100mmol/L Na<sub>2</sub>SO<sub>4</sub> + 0.05% NaN<sub>3</sub>
- Flow rate: 1.0mL/min
- Detection: UV@280nm
- Temperature: ambient
- Injection vol.: 10 $\mu$ L
- Samples: standard TSKgel SW<sub>XL</sub> test mixture:  
thyroglobulin (0.5g/L)  
 $\gamma$ -globulin (1g/L)  
ovalbumin (1g/L)  
ribonuclease A (1.5g/L)  
p-ABA (0.01g/L)



# Material and methods: Preparation of protein standards (desalting experiments)

<b>Protein</b>	<b>MW (kDa)</b>	<b>Concentration* (g/L approx.)</b>
ribonuclease A	14.7	19.5
thyroglobulin	670	11.3
$\gamma$ -globulin	150	14.5
ovalbumin	45	13.1
$\alpha$ -chymotrypsinogen	25.6	13.1
$\beta$ -lactoglobulin	18.4	10.8
lysozyme	14.7	11.6
myoglobin	16.7	14.5
cytochrome C	12.3	11.0
hemoglobin	68	11.9

\*in 100mmol/L phosphate buffer, pH 6.7

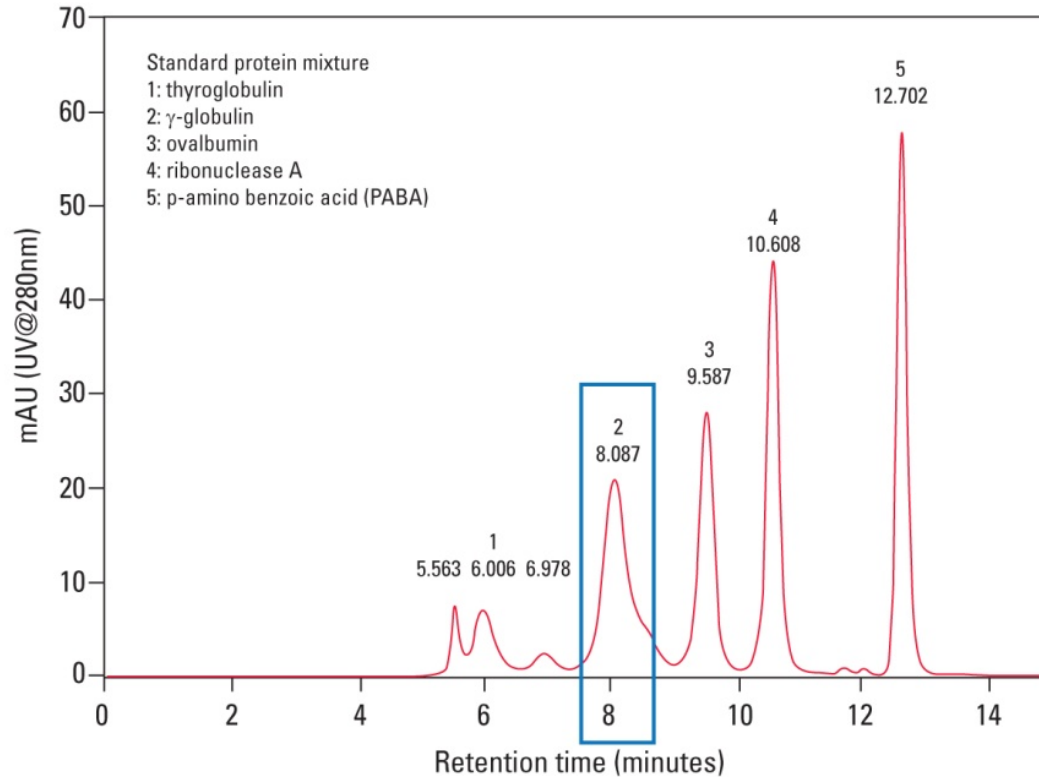


# Material and methods: Chromatographic conditions (desalting experiments)

- Columns : TSKgel BioAssist DS, 15 $\mu$ m, 4.6mm ID x 15cm, PEEK  
TSKgel BioAssist DS, 15 $\mu$ m, 10.0 mm ID x 15cm, PEEK
- Mobile Phase: 10mmol/L KH<sub>2</sub>PO<sub>4</sub>/K<sub>2</sub>HPO<sub>4</sub>, pH 6.7, 10mmol/L Na<sub>2</sub>SO<sub>4</sub> + 0.005% NaN<sub>3</sub>
- Flow rate: 0.8mL/min (4.6mm ID) and 1.0mL/min (10.0mm ID)
- Detection: UV@280nm and RI
- Temperature: ambient
- Injection vol.: 10 $\mu$ L unless mentioned otherwise
- Samples:  $\gamma$ -globulin was collected after injection of the standard TSKgel SW<sub>XL</sub> test mixture
- All analyses were carried out using an Agilent 1200 HPLC system run by Chemstation (ver B.04.01).
- All chemicals and standards were pure analytical grade from Sigma-Aldrich.
- Before injection, standards and samples were filtered through a 0.45 $\mu$ m filter.



# Separation of protein standard mixture using a TSKgel G3000SW<sub>XL</sub>, 5 $\mu$ m, 7.8mm ID $\times$ 30cm column

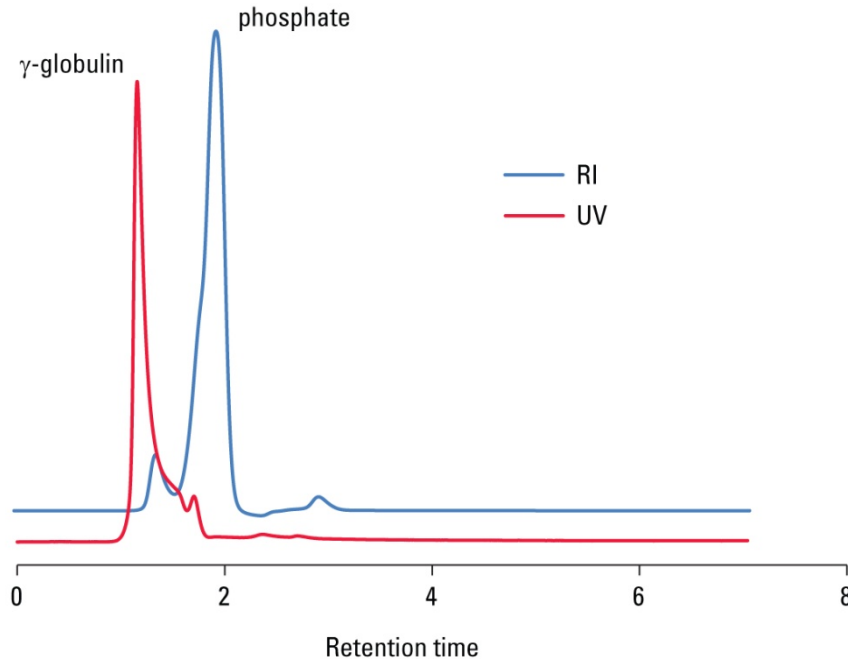


Mobile phase: 0.1mol/L  $\text{KH}_2\text{PO}_4/\text{Na}_2\text{HPO}_4$ , pH 6.7, 0.1mol/L  $\text{Na}_2\text{SO}_4$  + 0.05%  $\text{NaN}_3$

**10.0 $\mu$ L of  $\gamma$ -globulin (RT 8.087min) peak fraction was loaded into TSKgel BioAssist DS, 15 $\mu$ m, 4.6mm ID  $\times$  15cm column to desalt.**



# Desalting of $\gamma$ -globulin peak fraction using a TSKgel BioAssist DS, 15 $\mu$ m, 4.6mm ID $\times$ 15cm column

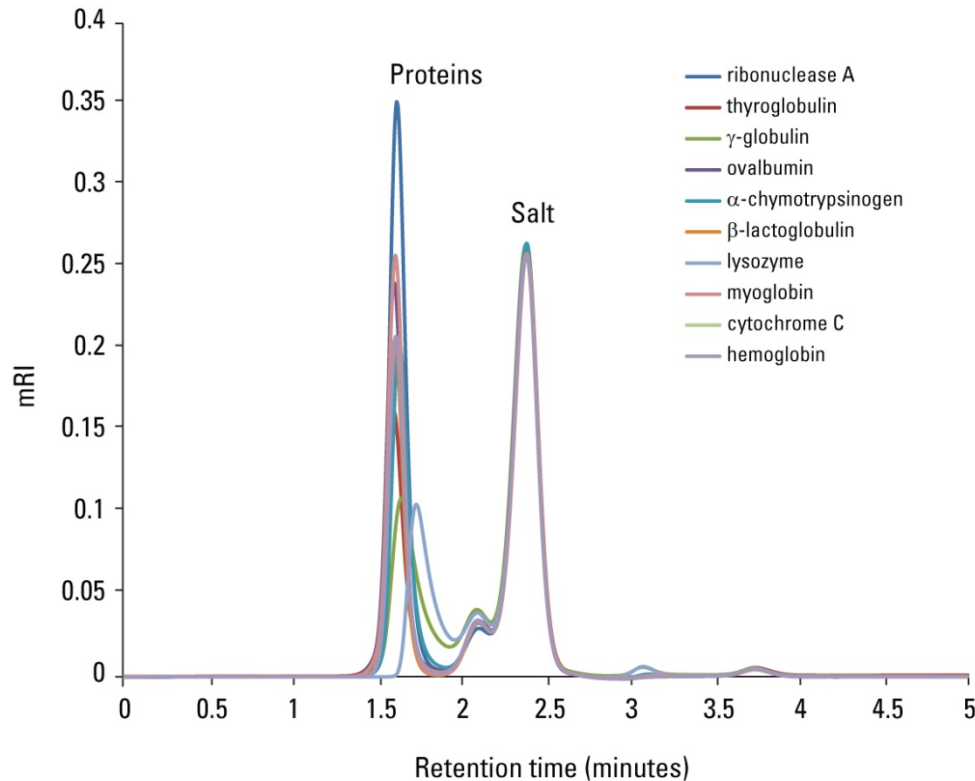


Mobile phase: 0.01mol/L  $\text{KH}_2\text{PO}_4/\text{NaHPO}_4$ , pH 6.7, 0.01mol/L  $\text{Na}_2\text{SO}_4$  + 0.05%  $\text{NaN}_3$   
Flow rate: 0.8mL/min.  
Detection: UV@280nm and RI  
Temperature: ambient

**Mobile phase  $\gamma$ -globulin fraction was efficiently desalted within a few minutes.**



# Desalting proteins using a TSKgel BioAssist DS, 15 $\mu$ m, 4.6mm ID x 15cm column



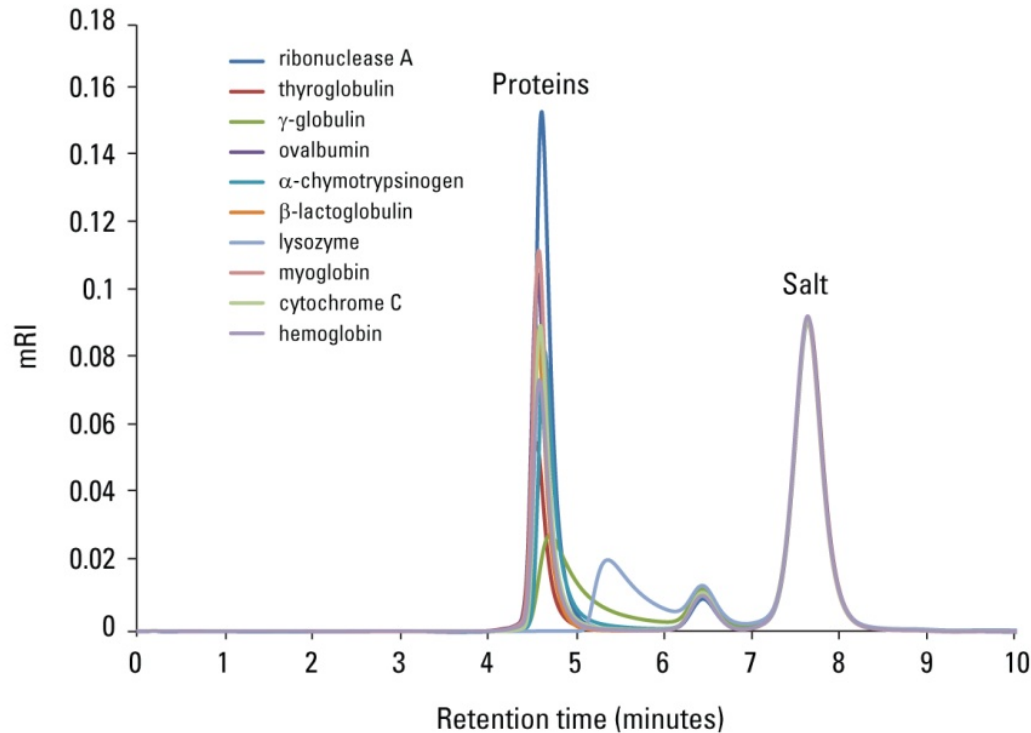
Proteins in 0.1mol/L phosphate buffer, pH 6.7

Mobile phase: 0.01mol/L  $\text{KH}_2\text{PO}_4/\text{Na}_2\text{HPO}_4$ , pH 6.7, 0.01mol/L  $\text{Na}_2\text{SO}_4$  + 0.05%  $\text{NaN}_3$

**Fast desalting with excellent reproducibility at analytical scale.**



# Desalting proteins using a TSKgel BioAssist DS, 15 $\mu$ m, 10mm ID x 15cm column



Proteins in 0.1mol/L phosphate buffer, pH 6.7

Mobile phase: 0.01mol/L  $\text{KH}_2\text{PO}_4/\text{Na}_2\text{HPO}_4$ , pH 6.7, 0.01mol/L  $\text{Na}_2\text{SO}_4$  + 0.05%  $\text{NaN}_3$

**Fast desalting with excellent reproducibility at semi-preparative scale.**



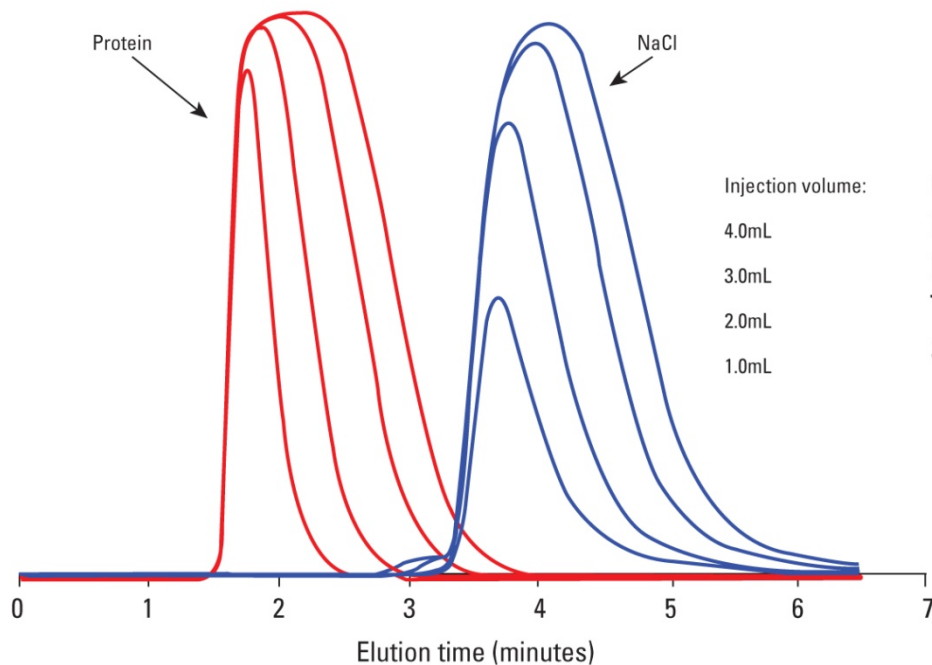
## Effect of sample load on efficiency of desalting of protein using TSKgel BioAssist DS, 15 $\mu$ m, 10.0mm ID x 15cm column

- The column has high loading (desalting) capacity.
- Less than 5% RSD (n=4) in efficiency up to a load of 1.5mg of Ribonuclease A .
- The resolution between the protein and salt peak was always >6.
- Even at ~2mg protein load of Ribonuclease A, the resolution between the protein and salt peak was 4.33.
- TSKgel BioAssist DS, 15 $\mu$ m, 4.6mm ID x 15cm column yielded a resolution of >2 at 1950 $\mu$ g load of Ribonuclease A (F=0.8mL/min).
- This study shows that both TSKgel BioAssist DS columns can be effectively used for desalting a large sample load.





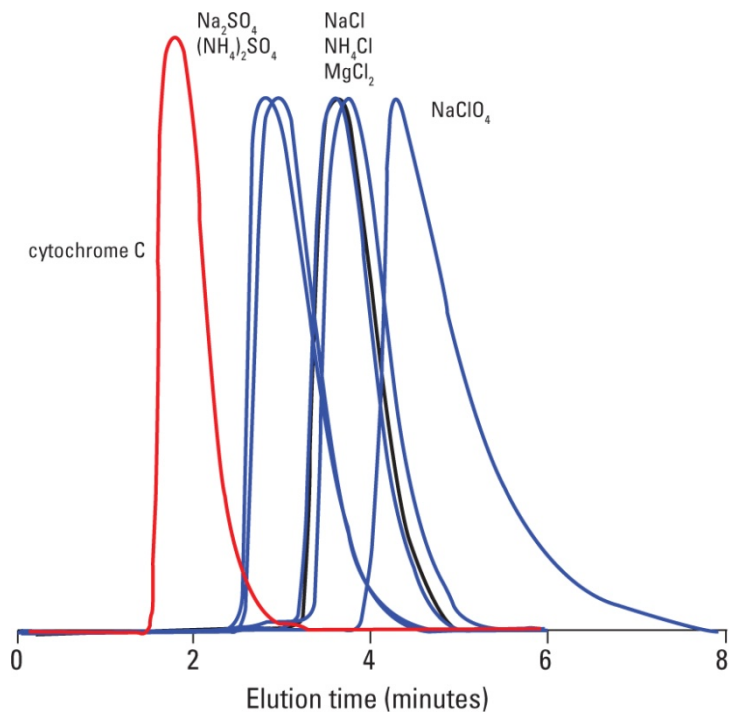
# Effect of injection volume on desalting profiles



Column: TSKgel BioAssist DS, 10mm ID x 15cm, PEEK  
Mobile phase: 50mmol/L sodium phosphate buffer, pH 6.5  
Flow rate: 3.0mL/min  
Temperature: 4°C  
Sample: 10mg/mL ovalbumin in 0.5mol/L NaCl and 25mmol/L sodium phosphate buffer, pH 6.5



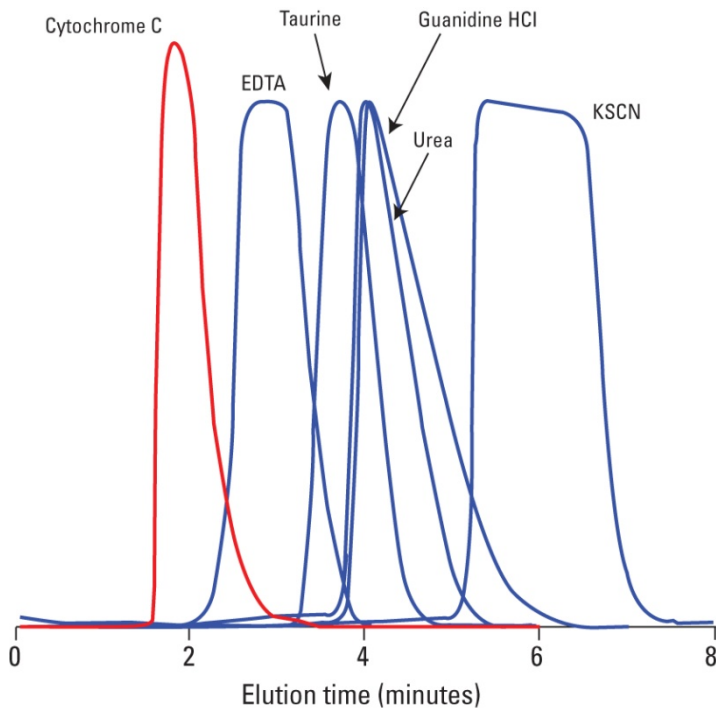
# Elution profiles of high salt concentration



Column: TSKgel BioAssist DS, 10mm ID x 15cm, PEEK  
Mobile phase: DI H<sub>2</sub>O  
Flow rate: 3.0mL/min  
Temperature: 4°C  
Injection vol.: 2mL  
Each salt concentration: 0.5mol/L



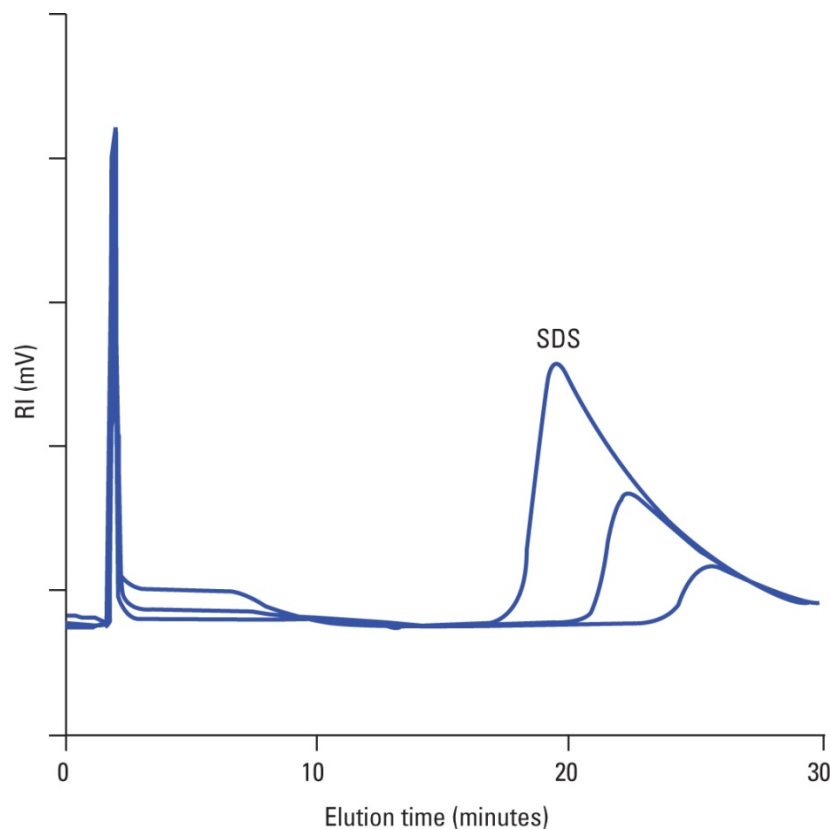
# Elution profiles of mobile phase additives



Column: TSKgel BioAssist DS, 10mm ID x 15cm, PEEK  
Mobile phase: DI H<sub>2</sub>O  
Flow rate: 3.0mL/min  
Temperature: 4°C  
Injection vol.: 2mL  
Each salt concentration: 0.5mol/L



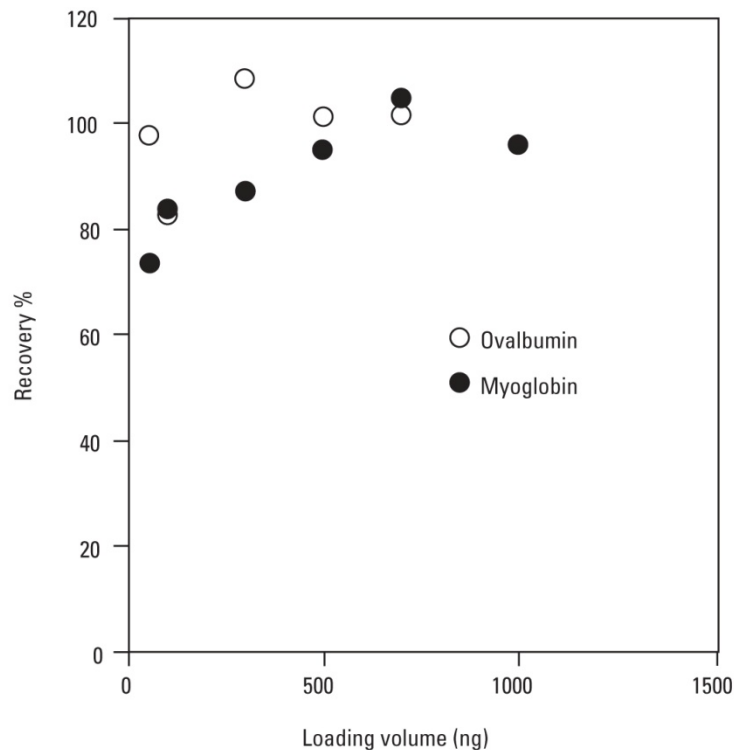
# Elution profiles of SDS



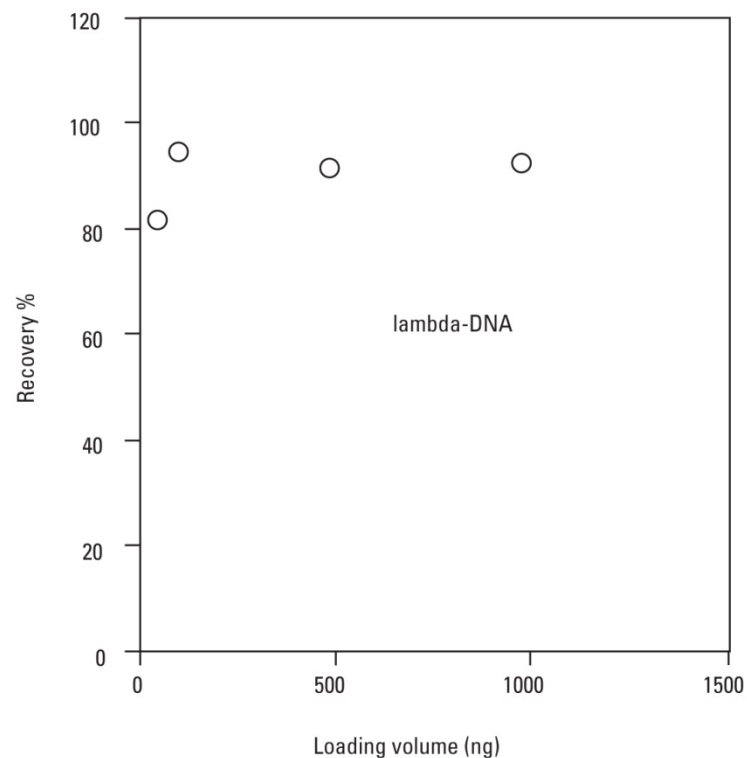
Column: TSKgel BioAssist DS,  
4.6mm ID x 15cm, PEEK  
Mobile phase: 20mmol/L phosphate buffer, pH 6.9  
Flow rate: 1.0mL/min  
Temperature: 25°C  
Sample: 0.1-0.5wt% SDS in eluent



# Recovery of selected proteins and DNA



Column: TSKgel BioAssist DS,  
4.6mm ID x 10cm, PEEK  
Mobile phase: 20mmol/L Tris-HCl buffer, pH 8.0  
for ovalbumin  
20mmol/L phosphate buffer, pH 6.5  
for myoglobin



Column: TSKgel BioAssist DS,  
4.6mm ID x 10cm, PEEK  
Mobile phase: 20mmol/L Tris-HCl buffer, pH 7.8



# Conclusions

- TSKgel BioAssist DS columns are designed for desalting of proteins and polynucleotides at semi-preparative scale with the following features:
  - 4-fold higher mechanical strength over that of conventional gels
  - Columns can be used at pressure up to 4MPa (600psi). Beads do not collapse at 12MPa pressure.
  - Exclusion limit of 2500Da (PEG)
  - Minimal secondary adsorption
  - Typical separation times of less than 5 minutes
  - High loading capacity
  - High recovery down to ng protein injected
  - Excellent reproducibility